

In Vitro Bioactivities of Alcohol-Free Benzydamine Oromucosal Solutions

Suwipa Ungphaiboon, Ph.D.¹, Sutasinee Ardhanwanich, M.Sc.¹, Sonsawan Kongpuckdee, Ph.D.², Duangkhae Maneenuan, B.Sc.¹, Teerapol Srichana, Ph.D.¹

¹Department of Pharmaceutical Technology and Drug Delivery System Excellence Center, Faculty of Pharmaceutical Sciences, Prince of Songkla University, Hat Yai, Songkhla 90110, Thailand.

²Department of Thai Traditional Medicine, Faculty of Health and Sports Science, Thaksin University, Pa Phayom, Phatthalung 93210, Thailand.

Received 14 July 2023 • Revised 20 July 2023 • Accepted 20 July 2023 • Published online 31 October 2023

Abstract:

Objective: This research aimed to evaluate the effects of chitosan and poloxamer 407 on *in vitro* cytotoxicity, anti-inflammatory, wound healing, and antimicrobial activities of 0.3% w/v benzydamine hydrochloride (HCl) oromucosal solutions.

Material and Methods: The effects of three alcohol-free benzydamine HCl oromucosal formulations; containing: 1) 0.5% w/v chitosan, 2) 15% w/v poloxamer 407, and 3) a combination of 0.5% w/v chitosan and 15% w/v poloxamer 407, on *in vitro* cytotoxicity and biological activities were evaluated and compared to a commercial benzydamine HCl mouth spray, containing alcohol and 0.12% chlorhexidine solution.

Results: All alcohol-free benzydamine HCl formulations and their vehicles were less cytotoxic to the macrophage RAW 264.7 cell line than the commercial spray and to the human gingival fibroblast cell line than chlorhexidine, respectively. The formulation containing 0.5% w/v chitosan exhibited the highest wound healing activity on the fibroblast cells among all tested products, and showed anti-inflammatory activity on macrophage cells, which is comparable to benzydamine HCl. Furthermore, from the time-kill assay, this formulation completely inhibited *Streptococcus mutans* within 1 minute, similar to 0.12% chlorhexidine solution, and reduced the cell number of *Candida albicans* more rapidly than the commercial spray.

Conclusion: The alcohol-free benzydamine HCl solution containing 0.5% w/v chitosan as a mucoadhesive polymer is a promising oral care candidate, which is safe to be used, and has wound healing, anti-inflammatory, and antimicrobial activities.

This paper was from The 7th Current Drug Development International Conference 2023 & The 1st World Kratom Conference (CDD2023 & WKC2023, August 22–25, 2023).

Contact: Suwipa Ungphaiboon, Ph.D.

Department of Pharmaceutical Technology, Faculty of Pharmaceutical Sciences,
Prince of Songkla University, Hat Yai, Songkhla 90110, Thailand.
E-mail: suwipa.u@psu.ac.th

J Health Sci Med Res
doi: 10.31584/jhsmr.20231002
www.jhsmr.org

© 2023 JHSMR. Hosted by Prince of Songkla University. All rights reserved.

This is an open access article under the CC BY-NC-ND license

(<http://www.jhsmr.org/index.php/jhsmr/about/editorialPolicies#openAccessPolicy>).

Keywords: alcohol-free, benzydamine, bioactivity, chitosan, poloxamer 407, oromucosal solution

Introduction

Oral mucositis is frequently observed in cancer patients undergoing chemotherapy, radiotherapy, or targeted therapy, causing cellular damage and ulceration¹. Several approaches and agents for preventing and treating oral mucositis are available, with variable efficacy. Oral mucositis can be managed by using artificial saliva, rinsing the mouth with saline, sodium bicarbonate, or special mouthwashes containing topical anesthetics (2% viscous lidocaine), mucosal coating agents, opioid analgesics; anti-inflammatory agents (benzydamine), antimicrobial agents, and antiseptic agents. Patients must avoid alcohol or alcohol-containing oral care products; additionally, Chlorhexidine mouthwash or other topical antimicrobials are not recommended for prophylaxis or treating oral mucositis caused by chemotherapy; except when oral hygiene is poor. On the other hand, benzydamine; a non-steroidal anti-inflammatory drug, has been recommended for patients with head and neck cancer undergoing radiation without chemotherapy; due to its anti-inflammatory, pain relieving, antipyretic, and antimicrobial activities¹⁻³.

Commercial benzydamine oromucosal solutions are available as mouthwashes and mouth sprays; however, there are some limitations due to a stinging sensation and short action. In addition, the pH of some commercial products is about 5, which promotes the erosion of teeth⁴. Moreover, formulations containing alcohol and strong flavoring agents can irritate and promote both dry mouth and inflammation, as the benzydamine is rapidly cleared from the mucosa by saliva flow. Benzydamine gel containing hydroxypropyl methyl cellulose has shown a statistically significant increase in the rate of oral mucosal repair in an animal model⁵. Therefore, alcohol-free benzydamine formulations with hydrating mucoadhesive polymers might be more appropriate to sustain the benzydamine in the oral cavity, as well as improving its pharmacological activities.

Chitosan is a cationic polysaccharide that possesses beneficial biological properties; such as biocompatibility, biodegradability, lack of allergenicity, non-toxicity, bioadhesion, hemostatic, wound healing, anti-inflammatory, antimicrobial, and moisturizing effect⁶. In addition, oral care products containing chitosan have been reported to prevent caries, control biofilm formation, and decrease pain and ulceration of stomatitis symptoms⁷⁻¹¹. Chitosan has also been shown to possess anti-inflammatory activity by reducing the interleukin-1 β -stimulated prostaglandin E2 protein levels in gingival fibroblasts¹². The combination of chitosan and other polymers; such as alginate, poloxamer, and polyethylene glycol, has been widely used in drug delivery systems to improve their properties and efficacy^{6,13-15}. Poloxamer 407, a non-ionic surfactant, is widely used in mucosal drug delivery systems due to its self-assembly into micelles, thermal reversibility, and biocompatibility. Although, poloxamer 407 lacks antibacterial properties, it has been shown to reduce the adhesion and biofilm formation of *Streptococcus mutans* (*S. mutans*) and *Candida* species as well as enhancing the rate of the wound healing process. An oral moisturizer containing 0.5% w/v chitosan and 15% w/v poloxamer 407 has been reported to demonstrate strong anti-adhesion and anti-biofilm properties against *Candida* species; however, it exhibited weaker anti-adhesion and anti-biofilm effects against *S. mutans* than the oral moisturizer containing only chitosan¹⁵.

Based on previous findings, this study was designed to evaluate the effects of alcohol-free benzydamine HCl oromucosal formulations containing chitosan and poloxamer 407 on *in vitro* cytotoxicity, anti-inflammatory, wound healing, and antimicrobial activities against *S. mutans* and *C. Albicans*, and compare them to a commercial 0.3% benzydamine HCl mouth spray and 0.12% chlorhexidine digluconate solution.

Material and Methods

Benzydamine HCl, chitosan (average viscosity molecular weight of 414 kDa; with a 91% degree of deacetylation), and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were purchased from Sigma-Aldrich, USA. Poloxamer 407 was obtained from Chanjao Longevity Company Limited, Thailand. Other excipients were pharmaceutical grade. The commercial benzydamine spray, composed of benzydamine HCl (3 mg/mL), ethanol, saccharin sodium, methyl hydroxybenzoate, glycerin, polyethylene glycol-40 hydrogenated castor oil, purified water, and a mint flavor, was used. Chlorhexidine digluconate was a gift from Songklanagarind Hospital. *S. mutans* DMST 18777 and *C. albicans* DMST 5815 were obtained from the culture collection of the Department of Medical Sciences, Ministry of Public Health, Thailand.

Alcohol-free benzydamine HCl oromucosal formulations

Three different oromucosal solutions of 0.3% w/v benzydamine HCl were formulated, by using 0.5% w/v chitosan, 15% w/v poloxamer 407, or a combination of 0.5% w/v chitosan and 15% w/v poloxamer 407; namely C0.5P0, C0P15, and C0.5P15, respectively. The compositions of alcohol-free benzydamine HCl oromucosal formulations and their physicochemical properties are shown in Table 1. The oromucosal formulations were prepared by mixing an aqueous solution of chitosan, pH 6–6.5, and an aqueous solution composed of benzydamine HCl, poloxamer 407, along with other additives, pH 6–6.5, and then by adding purified water to the required volume. The pH was measured using a digital pH meter (Mettler, Toledo, Switzerland); the viscosity was determined using a viscometer (Model DV-III Ultra, Brookfield, USA) at 25°C. All formulations were physicochemical stable after storage at 30°C and 45°C, with a relative humidity of 75% for 3 months. Vehicles composed of all ingredients; except benzydamine HCl, and

0.3% benzydamine HCl in water, were prepared as control formulations.

Anti-inflammatory activity studies and cytotoxicity in macrophage RAW 264.7 cell line

The inhibitory effect on nitric oxide (NO) production and cytotoxicity of samples using the macrophage RAW 264.7 cell line, a predominant producer of cytokines, were evaluated following the method of Tewtrakul et al. (2009)¹⁶. Briefly, the macrophage RAW 264.7 cells were seeded at a density of 1×10^5 cells/well in a 96-well plate (100 μ L). After 1 hr of incubation at 37°C in a humidified atmosphere containing 5% carbon dioxide (CO₂), the medium in a 96-well plate was replaced with a fresh medium containing of 0.2 μ g/mL of lipopolysaccharide (100 μ L), along with 100 μ L of samples in the ranges of concentrations at 1.7–13.6 mg/mL; equivalent to benzydamine HCl 5–40 μ g/mL. Then, the plates were incubated for 48 hr. The NO production was evaluated by measuring the accumulation of nitrite in the culture supernatant using the Griess reagent at a 1:1 ratio. The density of NO production was measured at 550 nm using a microplate reader. The anti-inflammatory activity of the developed formulations were compared to benzydamine HCl and a commercial benzydamine product. Caffeic acid phenethylester (nuclear factor kappa B inhibitor), at the range of concentration 0.5–5 μ g/mL and indomethacin (cyclooxygenase and inducible nitric oxide synthase inhibitor) at the range of concentration 1–40 μ g/mL, were used as standard positive controls. Cytotoxicity was determined using the MTT colorimetric method. The sample was considered cytotoxic if the density of the sample-treated group was less than 80% of the control group.

Wound healing by scratch test assay in a human gingival fibroblast (HGF) cell line

The scratch test assay was used to assess the wound healing activity of the sample, following the method

of Chaitrakoonthong et al. (2020).¹⁷ Briefly, HGF cells were seeded at a density of 1×10^5 cells/well in a 12-well plate and incubated for 24 hr at 37°C in a humidified atmosphere containing 5% CO₂ incubator to allow cell adhesion. A straight-line scratch was created in the native cell monolayer using a P1000 pipette tip. A fresh medium, or sample at the concentration of 0.34 mg/mL and 1.7 mg/mL, equivalent to benzydamine HCl 1 µg/mL and 5 µg/mL; respectively, was added to each well and incubated at 37°C. At 24, 48, and 72 hr, the wound area was observed and recorded using a phase-contrast inverted microscope. The remaining wound areas were analyzed using ImageJ software.

Antimicrobial and antibiofilm activities

Antimicrobial and antibiofilm activities against *S. mutans* DMST 18777 and *C. albicans* DMST 5815 of the alcohol-free benzydamine HCl formulations were determined in comparison with a commercial benzydamine spray and 0.12% w/v chlorhexidine solution.

The antimicrobial activity was evaluated using a colorimetric broth microdilution method, as described by Sungkharak et al. (2016).¹⁸ The final concentrations of 0.3% benzydamine samples were in the range of 4.9–2500 µg/mL, equivalent to benzydamine HCl of 1.5–750 µg/mL, and the final concentrations of vehicles and chitosan solution were in the range of 9.8–5000 µg/mL. The results were expressed as the minimum inhibitory concentration (MIC), minimum bactericidal concentration (MBC) and minimum fungicidal concentration (MFC) of the samples. The lowest concentration showing a blue color was considered as the MIC value based on the lack of reduction of the blue resazurin to the pink resorufin by the microbial dehydrogenase enzyme. The aliquots of broth from the wells containing no growth were inoculated by the streak plate method. The complete absence of growth was considered to represent the MBC or MFC.

The antibiofilm activities were determined using a microplate-based assay, as described by Perumal and

Mahmud (2013).¹⁹ The results were expressed as minimum biofilm inhibition concentration (MBIC), which is defined as the lowest concentration of an antimicrobial agent required to inhibit the formation of biofilms, The minimum biofilm eradication concentration (MBEC) can be defined as the lowest concentration of an antimicrobial agent required to eradicate biofilm. In addition, a time-kill assay was performed according to the method of Nittayananta et al. (2018).²⁰ The microbial suspensions were collected at different time points (1, 5, 15, 30, 60, 120, and 180 min) after exposure to the samples, and the reduction of microorganisms was evaluated using the standard plate count method.

Data analysis

All experiments were conducted in triplicate. The data are shown as the mean and standard deviation (S.D.). One-way analysis of variance (ANOVA) single factor was used to analyze the data, with the level of significance at a p-value less than 0.05.

Results

Anti-inflammatory activity and cytotoxicity in macrophage RAW 264.7 cell line

In vitro cytotoxicity testing of the formulations was conducted by measuring the cellular mitochondrial activity as a function of concentration. The commercial spray and the alcohol-free benzydamine HCl solutions exhibited no or negligible cytotoxicity (% relative cell viability greater than 80%) to the macrophage RAW 264.7 cell line at concentrations of less than 3.4 and 6.8 mg/mL, equivalent to benzydamine HCl of 10 and 20 µg/mL, respectively. However, the vehicles of the formulations and the positive controls exhibited no cytotoxicity at the highest concentration tested. The IC₅₀ value of C0.5P0 was comparable to that of benzydamine HCl and higher than those of commercial spray, C0P15 and C0.5P15; as shown in Table 2.

The anti-inflammatory activity of the oromucosal solutions was tested by measuring their inhibitory effect on NO production assay in the macrophage RAW 264.7 cell line. Caffeic acid phenethylester possessed the highest inhibitory effect on NO production, with an IC_{50} value of 0.91 ± 0.01 $\mu\text{g/mL}$, following that of the commercial spray; as shown in Table 2. The IC_{50} value of indomethacin was 14.88 ± 0.63 $\mu\text{g/mL}$, which was comparable to those of benzydamine HCl and the oromucosal solutions containing mucoadhesive polymers. However, the vehicles of the formulations had no anti-inflammatory activity.

Wound healing by scratch test assay in HGF cell line

Formulation C0.5P0, equivalent to benzydamine HCl 1 $\mu\text{g/mL}$, significantly accelerated the wound healing, as demonstrated by the proliferation and migration of HGF cells as well as complete wound closure after 72 hr of the scratch wound creator; as shown in Figure 1. Notably, HGF cells treated with C0.5P0 and its vehicle for 48 hr exhibited exceptionally high wound healing activity, with 95% and 100% closure, respectively. The alcohol-free benzydamine formulations showed a significant difference in cell migration to close the scratch wound compared to the control,

commercial spray, and chlorhexidine, while there were no differences between the formulations with mucoadhesive polymers and the benzydamine HCl in water. The wound healing activity of the solutions at a concentration equivalent to drug 5 $\mu\text{g/mL}$ was comparable to that of 1 $\mu\text{g/mL}$. However, chlorhexidine (5 $\mu\text{g/mL}$) was cytotoxic to HGF cells, and therefore no data of the percentage of wound healing is presented.

Antimicrobial and antibiofilm activities

The capability of C05P0 formulation to inhibit, kill, inhibit biofilm formation, and eradicate biofilm of *S. mutans* and *C. albicans* was in the range of concentration of benzydamine HCl and the commercial spray. However, the activities of benzydamine in various formulations were significantly lower than those of chlorhexidine; as shown in Table 3. Chitosan exhibited antimicrobial and antibiofilm activities against *S. mutans* and *C. albicans* at higher concentrations than benzydamine, while the vehicle had no such activities. There was no additive effect between benzydamine HCl and chitosan. From the time-kill assay, the C05P0 solution significantly decreased oral pathogens more rapidly than the commercial spray and C0.5P15; as shown in Figure 2.

Table 1 Composition and physicochemical properties of alcohol-free benzydamine HCl oromucosal formulations

Composition (%)	C0.5P0	C0P15	C0.5P15
Benzydamine HCl	0.3	0.3	0.3
Chitosan	0.5	0	0.5
Poloxamer 407	0	15	15
Sodium saccharin	0.05	0.05	0.05
Raspberry flavor	0.2	0.2	0.2
Methylparaben	0.1	0.1	0.1
Propylparaben	0.02	0.02	0.02
Propylene glycol	0.88	0.88	0.88
Lactic acid/ 1 N NaOH qs to pH 6–6.5	qs	qs	qs
Purified water qs to	100	100	100
Physicochemical properties			
Appearance	Clear solution	Clear solution	Clear solution
pH* (25°C)	6.21 ± 0.01	6.41 ± 0.00	6.51 ± 0.00
Viscosity* (25°C)	12.80 ± 0.35	123.37 ± 1.93	281.54 ± 1.83

*mean \pm S.D. (n=3), HCl=hydrochloride

Table 2 The anti-inflammatory activity and relative cell viability of benzydamine HCl oromucosal solutions tested by inhibitory effect on nitric oxide (NO) production assay in macrophage RAW 264.7 cell line as a measurement of anti-inflammatory activity

Sample	% NO inhibition: IC ₅₀ (µg/mL)*	% Relative cell viability: IC ₅₀ (µg/mL)*
Indomethacin	14.88±0.63 ^{b, c}	NA
Caffeic acid phenethylester	0.91±0.01 ^{a, c, d}	NA
Commercial spray ⁺	5.11±1.40 ^{a, b, d}	23.37±3.66 ^d
Benzydamine HCl	11.41±3.99 ^{b, c}	42.58±7.41 ^c
C0.5P0 ⁺	9.19±1.27 ^{a, b, c}	41.60±0.59 ^c
C0P15 ⁺	12.24±1.96 ^{b, c}	25.62±0.95 ^d
C0.5P15 ⁺	12.67±1.97 ^{b, c}	28.61±1.55 ^d

*mean±S.D., n=3, NA=not available

% NO inhibition=(A-B)/(A-C)×100, where;

A=the absorbance of lipopolysaccharide supernatant without a sample, B=the absorbance of lipopolysaccharide supernatant with sample, C=the absorbance of supernatant without lipopolysaccharide and sample; IC₅₀=the concentration that can inhibit NO cell secretion for 50%,⁺equivalent to benzydamine HCl.

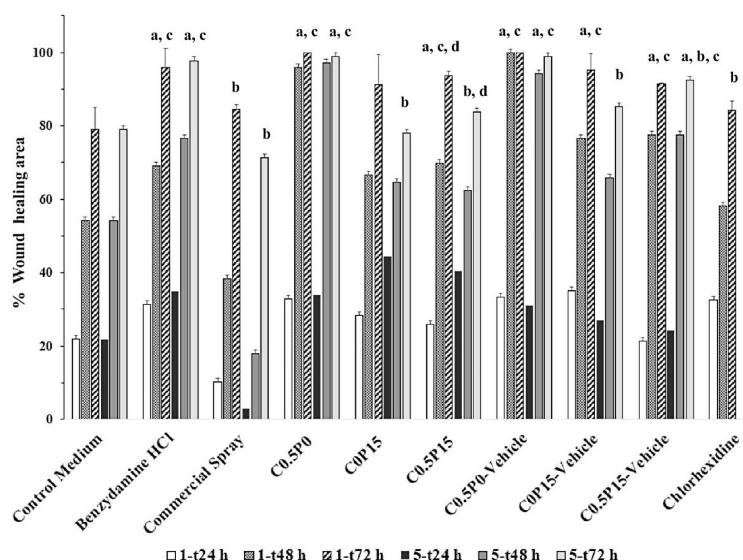
^{a, b, c, d}=p-value<0.05 when compared with indomethacin, caffeic acid phenethylester, commercial spray, and benzydamine HCl, respectively
HCl=hydrochloride

Table 3 The antimicrobial and antibiofilm activities of oromucosal solutions against *S. mutans* and *C. albicans*

Samples(µg/mL)	<i>S. mutans</i>				<i>C. albicans</i>			
	MIC	MBC	MBIC	MBEC	MIC	MFC	MBIC	MBEC
Chlorhexidine digluconate	0.98	1.95	1.95	7.81	15.63	31.25	15.63	62.50
Benzydamine HCl	188	188	188	375	375	375	188	750
Commercial Spray ⁺	375	750	375	375	375	750	94	>750
C0.5P0 ⁺	188	>750	94	375	375	375	375	750
C0.5P0-Vehicle	5000	NI	–	–	NI	–	–	–
Chitosan	1250	5000	1250	2500	>5000	–	>5000	>5000

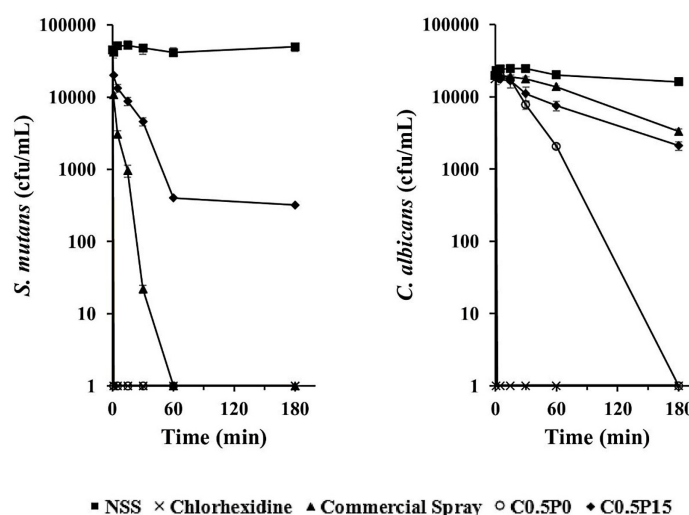
Data is expressed as mode (n=4), NI=Not inhibit, –=Not determined, ⁺equivalent to benzydamine HCl

S. mutans=*Streptococcus mutans*, *C. albicans*=*Candida albicans*, MIC=minimum inhibitory concentration, MBC=minimum bactericidal concentration, MBIC=minimum biofilm inhibitory concentration, MBEC=minimum biofilm eradication concentration, MFC= minimum fungicidal concentration, HCl=hydrochloride



mean±S.D., n=3 % Wound healing=[(Wound area of t0–Wound area of tn)/(Wound area of t0)]x100 Where;t0=Initial time of scratch test assay, tn=time of detection at 24, 48, or 72 hr.a, b, c, d=p-value<0.05 when compared with control medium, benzidine HCl, commercial spray, and vehicle, respectively.

Figure 1 The wound healing activity of alcohol-free benzidine HCl formulations and controls at the concentrations of 0.34 mg/mL and 1.70 mg/mL equivalent to benzidine HCl and chlorhexidine 1 µg/mL and 5 µg/mL in human gingival fibroblast cell line



(NSS=normal saline solution, chlorhexidine= 0.12% w/v chlorhexidine digluconate solution, and others=0.3% w/v benzidine HCl oromucosal solutions) (mean±S.D., n=3).

Figure 2 The reduction in cell number of *S. mutans* and *C. albicans* after exposure to oromucosal solutions for 180 min

Discussion

The prevention and management of oral mucositis are critical aspects of care for patients undergoing anticancer treatment. The severity of symptoms must be carefully considered to determine appropriate interventions. One strategy is modifying the patient's diet to avoid rough and sharp-edged foods; such as potato chips, as well as oral care products containing alcohol and strong flavoring agents, which may cause incidental trauma or burning. In this study, alcohol-free benzydamine solutions containing hydrating mucoadhesive polymers and mild flavoring agents were developed to sustain the drug on the oral mucosa and improve safety, wound healing, and antimicrobial activities.

Two mucoadhesive polymers, chitosan and poloxamer 407, were used to enhance the bioactivity of benzydamine in the treatment of oral mucositis. Mouthwash containing 0.5% w/v chitosan has been reported to be more effective than chlorhexidine in relieving oral mucositis symptoms¹⁰. Poloxamer 407 was used at 15% in the solution due to its thermo-responsive property.

The safety of oromucosal solutions was evaluated by their cytotoxicity to macrophage RAW 264.7 cells. The results indicated that all alcohol-free formulations showed cytotoxicity at a higher concentration compared to the commercial spray. Moreover, the vehicle of the formulations was found to be safer than benzydamine formulations. These findings suggest that the presence of alcohol and benzydamine HCl may increase the cytotoxicity of the product, while chitosan and poloxamer 407 may decrease it.

The anti-inflammatory activity of the oromucosal solutions was evaluated using an assay for inhibitory effect on NO production in macrophage RAW 264.7 cell line. The IC_{50} values of the oromucosal solutions containing mucoadhesive polymers were not significantly different from those of benzydamine HCl and indomethacin; however, they were approximately twice as high as those of the commercial spray. These results suggest that pharmaceutical excipients in the commercial spray may enhance the inhibitory effect

on NO; whereas, chitosan and poloxamer 407 do not affect the inhibitory effect of benzydamine.

The wound healing activity of the oromucosal solutions was evaluated in the HGF cell line. The results showed that wound healing did not improve when the concentration of benzydamine increased from 1 to 5 $\mu\text{g/mL}$. However, the solutions containing mucoadhesive polymers demonstrated a statistically significant increase in the rate of mucosal healing compared to the commercial spray. The percentage of wound healing area of the HGF cell line treated with the C0.5P0 solution (benzydamine HCl 1 $\mu\text{g/mL}$ and 5 $\mu\text{g/mL}$) and its vehicle was more than 90% after 48 hr of the scratch wound creator and closed to 100% after 72 hr. In contrast, the C0P15 solution accelerated wound repair less than the C0.5P0 solution, which may be due to the high viscosities preventing cell migration. These findings support previous studies indicating that chitosan promotes wound healing at different stages of the healing process by enhancing the functions of various inflammatory cells; including polymorphonuclear leukocytes, macrophages, and fibroblasts²¹. Furthermore, chitosan has been found to have good mucoadhesive properties, which can contribute to its effectiveness in promoting wound healing²². Additionally, chitosan has been shown to be an effective wound-healing accelerator when combined with other polymers²³.

The commercial spray exhibited the lowest wound healing activity among all tested solutions at 24 and 48 hr, which may be attributed to the excipients; such as alcohol and the mint flavor in the formulation. Alcohol is widely used in oral care products as a solvent for poorly soluble active ingredients and flavored oils, a penetration enhancer in addition to an active ingredient to control plaque-related oral disease. However, high concentrations of alcohol in the formulation may cause negative effects; such as burning sensations, pain, and dryness²⁴. Peppermint oil is popularly used in oral care products to reduce oral malodor and has anti-inflammatory, antimicrobial, and biofilm-inhibiting properties^{25,26}. Chlorhexidine at a concentration of 5 $\mu\text{g/L}$

mL induced a cytotoxic effect to HGF cells. Chlorhexidine mouthwash is commonly used to treat oral ulcers and prevent biofilm formation as well as target pathogenic microorganisms in the oral cavity. However, its long-term use may result in reversible discoloration of teeth and mucous membranes, altered taste perception, and serious side effects; such as mouth ulcers and swelling of the salivary glands²⁷. Therefore, the use of chlorhexidine at high concentrations and over prolonged periods must be monitored due to its potential toxicity and side effects.

C. albicans is an opportunistic pathogen that causes infections in immunosuppressed patients; such as cancer patients undergoing chemotherapy and elderly people with xerostomia, while *S. mutans* is the primary cause of dental caries. The MIC and MFC values of benzydamine against *C. albicans* have been reported to be 250 and 500 µg/mL, respectively²⁸. Various MIC values of different chitosans against *S. mutans* and *C. albicans* have been reported in a range of 0.08–5 mg/mL and 0.3–10 mg/mL, respectively^{8,29,30}. These differences have been attributed to their molecular weight, degree of deacetylation, pH of the growth medium, and the presence or absence of interfering substances. The antimicrobial action of chitosan has been proposed in different mechanisms. The interaction between positively charged chitosan molecules and negatively charged microbial cell membranes leads to the leakage of proteinaceous and other intracellular constituents as well as altering cell permeability³¹. In this study, the inhibitory activity and microbicidal effect values of chitosan and benzydamine were found to be similar to previous studies. However, there was no data on MBIC and MBEC of benzydamine and chitosan against *S. mutans* and *C. albicans*. The growth of *S. mutans* and *C. albicans* was completely inhibited by C0.5P0 within 1 min, which was as fast as that of a 0.12% w/v chlorhexidine solution, and 180 min, respectively, which was more rapid than with the commercial spray. The reduction in cell number of *S. mutans* and *C. albicans* by longer exposure times of C0.5P15 was found to correspond

to its thermo-responsive property. The antimicrobial activity of benzydamine was found to increase in the presence of chitosan, which is in agreement with previous studies²⁸.

Conclusion

In conclusion, the alcohol-free benzydamine HCl oromucosal solution containing 0.5% w/v chitosan and mild flavoring agents is a promising oromucosal candidate for the treatment of oral mucositis associated with anticancer therapy due to its safety, high wound healing rate, and significant antimicrobial activities.

One notable feature of this solution is its ability to promote wound healing, as evidenced by the complete closure of the scratch wound of the HGF at 72 hr. It was also found to be safer than a commercial spray that contains alcohol and a mint flavor, and a 0.12% chlorhexidine solution. Additionally, the solution demonstrated anti-inflammatory activity in the macrophage raw 264.7 cell line, which was comparable to that of benzydamine HCl. Furthermore, this formulation exhibited antimicrobial and antibiofilm activities against *S. mutans* and *C. albicans* and significantly inhibited the growth of oral pathogens faster than the commercial spray.

Acknowledgement

This study was funded by the Faculty of Pharmaceutical Sciences, Prince of Songkla University, Hat Yai, Thailand, (grant number PHA6404026S), and Prince of Songkla University Fundamental Fund (grant number PHA6505022M). We also acknowledge Dr. Saffanah Mohd Ab Azid for English editing.

Conflict of interest

There are no potential conflicts of interest to declare.

References

1. Brown TJ, Gupta A. Management of cancer therapy-associated oral mucositis. JCO Oncol Pract 2020;16:103–9.

2. Roopashri G, Jayanthi K, Guruprasad R. Efficacy of benzydamine hydrochloride, chlorhexidine, and povidone iodine in the treatment of oral mucositis among patients undergoing radiotherapy in head and neck malignancies: a drug trail. *Contemp Clin Dent* 2011;2:8–12.
3. Sheibani KM, Mafi AR, Moghaddam S, Taslimi F, Amiran A, Ameri A. Efficacy of benzydamine oral rinse in prevention and management of radiation-induced oral mucositis: A double-blind placebo-controlled randomized clinical trial. *Asia Pac J Clin Oncol* 2015;11:22–7.
4. Vivek S, Shwetha R. Endogenous pH, titratable acidity of commercially available mouthwashes in Indian market. *Int J Clin Trials* 2015;2:20–4.
5. Karavana (Hızarcıoğlu) SY, Sezer B, Güneri P, Veral A, Boyacıoğlu H, Ertan G, et al. Efficacy of topical benzydamine hydrochloride gel on oral mucosal ulcers: an in vivo animal study. *Int J Oral Maxillofac Surg* 2011;40:973–8.
6. Fatullayeva S, Tagiyev D, Zeynalov N, Mammadova S, Aliyeva E. Recent advances of chitosan-based polymers in biomedical applications and environmental protection. *J Polym Res* 2022;29:259.
7. Carlson RP, Taffs R, Davison WM, Stewart PS. Anti-biofilm properties of chitosan-coated surfaces. *J Biomater Sci Polym Ed* 2008;19:1035–46.
8. Costa E, Silva S, Tavaría F, Pintado M. Antimicrobial and antibiofilm activity of chitosan on the oral pathogen *Candida albicans*. *Pathogens* 2014;3:908–19.
9. Mahattanadul S, Mustafa MW, Kuadkaew S, Pattharachayakul S, Ungphaiboon S, Sawanyawisuth K. Oral ulcer healing and anti-Candida efficacy of an alcohol-free chitosan-curcumin mouthwash. *Eur Rev Med Pharmacol Sci* 2018;22:7020–3.
10. Mahima VG, Patil K, Kulkarni PK, Tayal S, Keshari D. Use of chitosan mouth-wash in radio-chemotherapy induced oral mucositis: A case-control study. *J Adv Clin Res Insights* 2015;2:248–52.
11. Pasquantonio G, Greco C, Prenna M, Ripa C, Vitali LA, Petrelli D, et al. Antibacterial activity and anti-biofilm effect of chitosan against strains of *Streptococcus mutans* isolated in dental plaque. *Int J Immunopathol Pharmacol* 2008;21:993–7.
12. Arancibia R, Maturana C, Silva D, Tobar N, Tapia C, Salazar JC, et al. Effects of chitosan particles in periodontal pathogens and gingival fibroblasts. *J Dent Res* 2013;92:740–5.
13. Paul W, Sharma C, Tirunal C. Chitosan and alginate wound dressings: a short review. *Trends in biomaterials & artificial organs* [serial on the Internet]. 2004 [cited 2023 May 2]; Available from: <https://www.semanticscholar.org/paper/Chitosan-and-Alginate-Wound-Dressings%3A-A-Short-Paul-Sharma/b38c2de711be7a10a802c8534a7879b6801186ef>
14. Bansal M, Mittal N, Yadav SK, Khan G, Gupta P, Mishra B, et al. Periodontal thermoresponsive, mucoadhesive dual antimicrobial loaded in-situ gel for the treatment of periodontal disease: Preparation, in-vitro characterization and antimicrobial study. *J Oral Biol Craniofac Res* 2018;8:126–33.
15. Sarideechaigul W, Ungphaiboon S, Ardpolthai H, Jehsama-ae A, Sangsuttiwongsa K, Taweechaisupapong S. Effect of oral moisturizers containing chitosan and poloxamer 407 on biofilm formation of *Candida* species and *Streptococcus mutans*: in vitro. *Songklanakarin J Sci Technol* 2022;44:1298–305.
16. Tewtrakul S, Tansakul P, Panichayupakaranant P. Effects of rhinacanthins from *Rhinacanthus nasutus* on nitric oxide, prostaglandin E2 and tumor necrosis factor- α releases using RAW264.7 macrophage cells. *Phytomedicine* 2009;16:581–5.
17. Chaitrakoonthong T, Ampornaramveth R, Kamolratanakul P. Rinsing with l-ascorbic acid exhibits concentration-dependent effects on human gingival fibroblast in vitro wound healing behavior. *Int J Dent* 2020;2020:4706418.
18. Sungkharak S, Ungphaiboon S. Antibacterial activity against acne involved bacteria of chitosan in a soluble state and as nanoparticles. *Chiang Mai J Sci* 2016;43:1150–9.
19. Perumal S, Mahmud R. Chemical analysis, inhibition of biofilm formation and biofilm eradication potential of *Euphorbia hirta* L. against clinical isolates and standard strains. *BMC Complement Altern Med* 2013;13:346.
20. Nittayananta W, Limsuwan S, Srichana T, Sae-Wong C, Amnuait T. Oral spray containing plant-derived compounds is effective against common oral pathogens. *Arch. Oral Biol* 2018;90:80–5.
21. Dai T, Tanaka M, Huang YY, Hamblin MR. Chitosan preparations for wounds and burns: antimicrobial and wound-healing effects. *Expert Rev Anti Infect Ther* 2011;9:857–79.
22. Hurler J, Skalko-Basnet N. Potentials of chitosan-based delivery systems in wound therapy: bioadhesion study. *J Funct Biomater* 2012;3:37–48.
23. Liu H, Wang C, Li C, Qin Y, Wang Z, Yang F, et al. A functional

- chitosan-based hydrogel as a wound dressing and drug delivery system in the treatment of wound healing. RSC Adv 2018;8:7533–49.
24. Werner CW de A, Seymour RA. Are alcohol containing mouthwashes safe? Br Dent J 2009;207:E19.
25. Haghgoo R, Abbasi F. Evaluation of the use of a peppermint mouth rinse for halitosis by girls studying in Tehran high schools. J Int Soc Prev Community Dent 2013;3:29–31.
26. Radu CM, Radu CC, Bochiş SA, Arbănaşi EM, Lucan AI, Murvai VR, et al. Revisiting the therapeutic effects of essential oils on the oral microbiome. Pharmacy 2023;11:33.
27. Prasanna SGV, Lakshmanan DR. Characteristics, uses and side effects of chlorhexidine—A review. IOSR J Dent Med Sci 2016;15:57–9.
28. Rossi S, Marciello M, Bonferoni MC, Ferrari F, Sandri G, Dacarro C, et al. Thermally sensitive gels based on chitosan derivatives for the treatment of oral mucositis. Eur J Pharm Biopharm 2010;74:248–54.
29. Aranaz I, Acosta N, Civera C, Elorza B, Mingo J, Castro C, et al. Cosmetics and cosmeceutical applications of chitin, chitosan and their derivatives. Polymers 2018;10:213.
30. Gomaa EZ. Improvement of antimicrobial and anti-biofilm potentials of mouthwashes by chitosan produced by lactic acid bacteria: an *in vitro* study. J Microbiol Biotechnol 2017;2:52–7.
31. Rabea EI, Badawy MET, Stevens CV, Smagghe G, Steurbaut W. Chitosan as antimicrobial agent: applications and mode of action. Biomacromolecules 2003;4:1457–65.